

RESEARCH PROGRESS ON LONG NON-CODING RNA-MEDIATED SEPSIS PROGRESSION

HongYan Ren^{1,2}, JianQuan Li^{2*}

¹Medical School, Guizhou University, Guiyang 550000, Guizhou, China.

²Intensive Care Unit, Guizhou Provincial People's Hospital, Guiyang 550000, Guizhou, China.

Corresponding Author: Jianquan Li, Email: lijianquan7205@163.com

Abstract: Sepsis represents a severe dysregulation of the host response to infection. In recent years, the involvement of long non-coding RNAs (lncRNAs) in the onset and progression of sepsis has garnered significant attention. This review outlines the expression profiles and regulatory functions of lncRNAs in sepsis, emphasizing their associations with inflammatory processes, immune dysregulation, and organ injury. It highlights the molecular pathways through which lncRNAs influence sepsis by modulating inflammatory signaling, immune cell activities, and programmed cell death. Additionally, the potential utility of lncRNAs as diagnostic markers and therapeutic targets is discussed.

Keywords: Long non-coding RNA; Sepsis; Inflammatory response; Immune regulation; Organ injury; Molecular mechanism

1 INTRODUCTION

Sepsis remains a major contributor to global mortality, characterized by a complex pathophysiology involving uncontrolled inflammation, immune impairment, and multi-organ failure [1]. Although treatment strategies have advanced, mortality rates persist at high levels, underscoring the need for novel diagnostic and therapeutic approaches. Long non-coding RNAs (lncRNAs), which exceed 200 nucleotides and lack protein-coding potential, have emerged as crucial regulators in numerous diseases. Research indicates that lncRNAs participate in gene expression control, chromatin organization, and protein interactions, impacting key processes such as inflammation, immunity, and cell death [2-5]. Advances in high-throughput sequencing have unveiled dysregulated lncRNA expression in sepsis, shedding light on their functional importance. This article synthesizes current knowledge on lncRNA mechanisms in sepsis and explores their clinical potential.

2 OVERVIEW OF LONG NON-CODING RNAs

lncRNAs are RNA molecules longer than 200 nucleotides with limited protein-coding capability. They are categorized based on genomic context into sense, antisense, bidirectional, intronic, and intergenic types. Unlike shorter non-coding RNAs like miRNAs, lncRNAs exhibit complex structures and operate through diverse mechanisms [6-7].

The functional spectrum of lncRNAs includes: (1) serving as scaffolds in ribonucleoprotein complexes; (2) acting as competing endogenous RNAs (ceRNAs) to sequester miRNAs; (3) guiding chromatin-modifying complexes; (4) influencing transcriptional activity; (5) modulating mRNA stability and translation; and (6) facilitating intercellular communication. These roles position lncRNAs as pivotal regulators within gene networks, contributing to both normal physiology and disease [8-10].

3 EXPRESSION PATTERNS OF LONG NON-CODING RNAs IN SEPSIS

High-throughput studies have identified extensive alterations in lncRNA expression in sepsis patients and experimental models [11]. For instance, peripheral blood mononuclear cells from septic individuals show hundreds of differentially expressed lncRNAs, some correlating with disease severity and outcomes [12]. Upregulated lncRNAs such as NEAT1, MALAT1, and HOTAIR, and downregulated ones like MEG3 and GAS5, are frequently reported [13-15].

Organ-specific lncRNA expression changes are also evident. In septic acute lung injury, PFI and lnc-IL7R are elevated [16-17]; in kidney injury, TUG1 and AK139328 are altered [18]; and in myocardial dysfunction, H19 and KCNQ1OT1 are dysregulated [19-20]. These variations suggest tissue-specific regulatory roles. Moreover, lncRNA expression dynamically shifts with sepsis progression: pro-inflammatory types like THRIL and lncRNA-ERS rise during hyperinflammation, while immunomodulatory lncRNAs such as NR_045064 and uc.48+ dominate during immunosuppression.

4 MECHANISMS OF LNCRNAs IN SEPSIS PATHOGENESIS

4.1 lncRNAs and Inflammatory Response in Sepsis

LncRNAs modulate sepsis-associated inflammation through various avenues. As ceRNAs, they sequester miRNAs to derepress inflammatory mediators. For example, NEAT1 sponges miR-125a-5p, alleviating suppression of TRAF6 and activating NF- κ B to enhance cytokine production [21-22]. Similarly, MALAT1 influences the miR-146a/TRAF6 axis, impacting TLR4 signaling [23-24].

Direct involvement in inflammatory pathways is also common. THRIL complexes with hnRNPL to modulate TNF- α transcription [25-26], while LincRNA-EPS interacts with hnRNPL to suppress inflammatory genes [27]. The pseudogene-derived lncRNA Lethe binds NF- κ B, curtailing its activity [26, 28-29]. These interactions fine-tune inflammatory responses.

LncRNAs also regulate inflammasomes: MEG3 inhibits NLRP3 assembly [30-31], and GAS5 binds glucocorticoid receptors to boost anti-inflammatory cytokines like IL-10 [32]. Conversely, SNHG1 promotes NLRP3 activation, worsening inflammation [33].

4.2 lncRNAs and Immune Regulation in Sepsis

4.2.1 Immune cell modulation by lncRNAs

Immune cell functions are extensively regulated by lncRNAs. In macrophages, TUG1 upregulation upon LPS stimulation enhances activation and cytokine release; its knockdown mitigates these effects [34]. Mechanistically, TUG1 binds miR-142-3p to elevate NCOA1, influencing polarization and inflammation [35]. In lymphocytes, NRON dysregulation in septic T cells impairs calcineurin-NFAT signaling, leading to immune dysfunction [36-37].

4.2.2 lncRNAs and immune evasion

Pathogens exploit lncRNAs to evade immunity. Viral lncRNAs, like HCMV's β 2.7, bind hnRNP A2/B1 to suppress interferon responses, aiding persistence [38-40]. Bacterial infections may similarly alter host lncRNAs to impair immunity, though this area requires further study [41-42].

5 LNCRNAs IN CELL DEATH AND ORGAN DAMAGE IN SEPSIS

5.1 lncRNAs and Apoptosis

Apoptosis of immune and parenchymal cells contributes significantly to sepsis pathology. LncRNA-ATB upregulation in septic T cells promotes apoptosis via miR-200c/ZEB1 signaling [43-47]. In macrophages, MEG3 overexpression accelerates apoptosis through p53 activation [48-49].

In organ cells, HOTTIP upregulation exacerbates cardiomyocyte apoptosis via miR-125a-5p/Bax [50-51], while MIAT sponges miR-205 to increase Caspase-3 expression, worsening renal injury [52-54].

5.2 lncRNAs and Tissue/Organ Injury

LncRNAs are implicated in organ-specific damage: PFI and MALAT1 aggravate lung injury by promoting apoptosis and endothelial permeability [55-56], whereas MEG3 is protective [31]. In the kidney, TUG1 and H19 exacerbate injury through HMGB1 and let-7/STAT3 pathways [57-58]. In the heart, H19 and KCNQ1OT1 affect contractility and apoptosis [59-60].

6 LNCRNAs AND COAGULATION ABNORMALITIES

Coagulopathy is a critical aspect of sepsis. UCA1 upregulation in sepsis enhances coagulation via miR-143-3p/TF signaling [61-62]. LncRNAs may also influence platelet function, though detailed mechanisms remain elusive.

7 DIAGNOSTIC AND THERAPEUTIC POTENTIAL OF LNCRNAs IN SEPSIS

LncRNAs show promise as biomarkers: panels like MALAT1/HOTAIR/GAS5 improve diagnostic accuracy and prognosis prediction [63]. Therapeutically, targeting pro-inflammatory lncRNAs (e.g., NEAT1, THRIL) or supplementing protective ones (e.g., MEG3, GAS5) ameliorates sepsis in models [64]. Nanocarrier systems may enhance delivery, but challenges in specificity and safety remain.

8 CONCLUSIONS AND PERSPECTIVES

This review outlines the expanding roles of lncRNAs in sepsis pathophysiology. Their involvement in inflammation, immunity, and organ injury provides insights into disease mechanisms and highlights translational opportunities. Outstanding issues include: (1) validating lncRNA biomarkers in larger cohorts; (2) elucidating spatiotemporal regulation; (3) exploring interactions with other epigenetic mechanisms; and (4) improving therapeutic delivery. Future directions involve standardizing detection methods, integrating multi-omics data, understanding heterogeneity, and advancing precision medicine. LncRNAs hold potential as novel targets for improving sepsis outcomes.

COMPETING INTERESTS

The authors have no relevant financial or non-financial interests to disclose.

FUNDING

This work was funded by Foundation of Guizhou Science and Technology Cooperation -zk[2023] General 228 and Science and Technology Fund of Guizhou Provincial Health and Wellness Committee in 2023 (gzwkj2023-008).

REFERENCES

- [1] Cao M, Wang G, Xie J. Immune dysregulation in sepsis: experiences, lessons and perspectives. *Cell Death Discovery*, 2023, 9(1): 465.
- [2] Zhang D, Pei S, Feng Z, et al. Functions and mechanisms of lncRNAs in immune escape and their application in immunotherapy for colorectal cancer. *Journal of Translational Medicine*, 2025, 23(1): 689.
- [3] Zhang Y, Liu H, Niu M, et al. Roles of long noncoding RNAs in human inflammatory diseases. *Cell Death Discovery*, 2024, 10(1): 235.
- [4] Jin L, Liao J, Jin L, et al. Critical role of lncRNA in sepsis-associated acute kidney injury. *Frontiers in Pharmacology*, 2025, 16: 1627253.
- [5] Leng L, Wang H, Hu Y, et al. LINC02363: a potential biomarker for early diagnosis and treatment of sepsis. *BMC Immunology*, 2025, 26(1): 23.
- [6] Ma L, Bajic VB, Zhang Z. On the classification of long non-coding RNAs. *RNA Biology*, 2013, 10(6): 925-33.
- [7] Chodurska B, Kunej T. Long non-coding RNAs in humans: Classification, genomic organization and function. *Non-coding RNA Research*, 2025, 11: 313-327.
- [8] Gao N, Li Y, Li J, et al. Long Non-Coding RNAs: The Regulatory Mechanisms, Research Strategies, and Future Directions in Cancers. *Front Oncology*, 2020, 10: 598817.
- [9] Tang J., Zhang J, Lu Y, et al. Novel insights into the multifaceted roles of m6A-modified lncRNAs in cancers: biological functions and therapeutic applications. *Biomarker Research*, 2023, 11(1): 42.
- [10] Zhang X, Wang W, Zhu W, et al. Mechanisms and Functions of Long Non-Coding RNAs at Multiple Regulatory Levels. *International Journal of Molecular Sciences*, 2019, 20(22).
- [11] Shin JJ, Park J, Shin H, et al. Roles of lncRNAs in NF- κ B-Mediated Macrophage Inflammation and Their Implications in the Pathogenesis of Human Diseases. *International Journal of Molecular Sciences*, 2024. 25(5): 2670.
- [12] Zhang W, Li Y, Li G, et al. Identification of lncRNAs in peripheral blood mononuclear cells associated with sepsis immunosuppression based on weighted gene co-expression network analysis. *Hereditas*, 2025, 162(1): 51.
- [13] Wen M, Cai G, Ye J, et al. Single-cell transcriptomics reveals the alteration of peripheral blood mononuclear cells driven by sepsis. *Annals of Translational Medicine*, 2020, 8(4): 125.
- [14] Wang W, Yang N, Wen R, et al. Long Noncoding RNA: Regulatory Mechanisms and Therapeutic Potential in Sepsis. *Frontiers in Cellular and Infection Microbiology*, 2021, 11: 563126.
- [15] Fan XY, Ma ZX, Tang LB, et al. lncRNA NEAT1 mediates LPS-induced pyroptosis of BEAS-2B cells via targeting miR-26a-5p/ROCK1 axis. *Kaohsiung Journal of Medical Sciences*, 2023, 39(7): 665-674.
- [16] Cui H, Xie N, Tan Z, et al. The human long noncoding RNA lnc-IL7R regulates the inflammatory response. *European Journal of Immunology*, 2014, 44(7): 2085-95.
- [17] Sun J, Jin T, Su W, et al. The long non-coding RNA PFI protects against pulmonary fibrosis by interacting with splicing regulator SRSF1. *Cell Death & Differentiation*, 2021, 28(10): 2916-2930.
- [18] Chen T, Lu J, Fan Q. lncRNA TUG1 and kidney diseases. *BMC Nephrology*, 2025, 26(1): 139.
- [19] Sun F, Yuan W, Wu H, et al. lncRNA KCNQ1OT1 attenuates sepsis-induced myocardial injury via regulating miR-192-5p/XIAP axis. *Experimental Biology and Medicine*, 2020, 245(7): 620-630.
- [20] Fang Y, Hu J, Wang Z, et al. lncRNA H19 functions as an Aquaporin 1 competitive endogenous RNA to regulate microRNA-874 expression in LPS sepsis. *Biomedicine & Pharmacotherapy*, 2018, 105: 1183-1191.
- [21] Gan Y, Long J, Zeng Y, et al. lncRNA IL-17RA-1 Attenuates LPS-Induced Sepsis via miR-7847-3p/PRKCG-Mediated MAPK Signaling Pathway. *Mediators of Inflammation*, 2022, 2022: 9923204.
- [22] Feng F, Jiao P, Wang J, et al. Role of Long Noncoding RNAs in the Regulation of Cellular Immune Response and Inflammatory Diseases. *Cells*, 2022, 11(22): 3642.
- [23] Liao Z, Zheng R, Shao G. Mechanisms and application strategies of miRNA-146a regulating inflammation and fibrosis at molecular and cellular levels (Review). *International Journal of Molecular Medicine*, 2023, 51(1).
- [24] Huang G, Zhao X, Bai Y, et al. Regulation of mitochondrial autophagy by lncRNA MALAT1 in sepsis-induced myocardial injury. *European Journal of Medical Research*, 2024, 29(1): 524.
- [25] Li Z, Chao T, Chang K, et al. The long noncoding RNA THRIL regulates TNF α expression through its interaction with hnRNPL. *Proceedings of the National Academy of Sciences of the United States of America*, 2014, 111(3): 1002-7.
- [26] Rapicavoli NA, Qu K, Zhang J, et al. A mammalian pseudogene lncRNA at the interface of inflammation and anti-inflammatory therapeutics. *eLife*, 2013, 2: e00762.

- [27] Atianand MK, Hu W, Satpathy AT, et al. A Long Noncoding RNA lincRNA-EPS Acts as a Transcriptional Brake to Restrain Inflammation. *Cell*, 2016, 165(7): 1672-1685.
- [28] Gupta SC, Awasthee N, Rai V, et al. Long non-coding RNAs and nuclear factor- κ B crosstalk in cancer and other human diseases. *Biochimica et Biophysica Acta - Reviews on Cancer*, 2020, 1873(1): 188316.
- [29] Statello L, Guo C, Chen L, et al. Gene regulation by long non-coding RNAs and its biological functions. *Nature Reviews Molecular Cell Biology*, 2021, 22(2): 96-118.
- [30] Gao H, Zhang X, Tang F, et al. Knockdown of lncRNA MEG3 protects against sepsis-induced acute lung injury in mice through miR-93-5p-dependent inhibition of NF- κ B signaling pathway. *Pathology Research and Practice*, 2022, 239: 154142.
- [31] Liao H, Zhang S, Qiao J. Silencing of long non-coding RNA MEG3 alleviates lipopolysaccharide-induced acute lung injury by acting as a molecular sponge of microRNA-7b to modulate NLRP3. *Aging (Albany NY)*, 2020, 12(20): 20198-20211.
- [32] Keenan CR, Schuliga MJ, Stewart AG. Pro-inflammatory mediators increase levels of the noncoding RNA GAS5 in airway smooth muscle and epithelial cells. *Canadian Journal of Physiology and Pharmacology*, 2015, 93(3): 203-206.
- [33] Cao B, Wang T, Qu Q, et al. Long Noncoding RNA SNHG1 Promotes Neuroinflammation in Parkinson's Disease via Regulating miR-7/NLRP3 Pathway. *Neuroscience*, 2018, 388: 118-127.
- [34] Ma W, Zhang W, Cui B, et al. Functional delivery of lncRNA TUG1 by endothelial progenitor cells derived extracellular vesicles confers anti-inflammatory macrophage polarization in sepsis via impairing miR-9-5p-targeted SIRT1 inhibition. *Cell Death & Disease*, 2021, 12: 1056.
- [35] Zhang R, Huang X, Jiang Y, et al. LncRNA TUG1 regulates autophagy-mediated endothelial-mesenchymal transition of liver sinusoidal endothelial cells by sponging miR-142-3p. *American Journal of Translational Research*, 2020, 12(3): 758-772.
- [36] Ahmad I, Valverde A, Ahmad H, et al. Long Noncoding RNA in Myeloid and Lymphoid Cell Differentiation, Polarization and Function. *Cells*, 2020, 9(2): 269.
- [37] Yao Z, Xiong Z, Li R, et al. Long non-coding RNA NRON is downregulated in HCC and suppresses tumour cell proliferation and metastasis. *Biomedicine & Pharmacotherapy*, 2018, 104: 102-109.
- [38] Boliar S, Prats-Mari L, Fortes P. Editorial: Long non-coding RNAs in viral infections and immunity. *Frontiers in Immunology*, 2023(14).
- [39] Zhang D, Zhang M, Zhang L, et al. Long non-coding RNAs and immune cells: Unveiling the role in viral infections. *Biomedicine & Pharmacotherapy*, 2024, 170: 115978.
- [40] Perera MR, Sinclair JH, Sinclair JH. The Human Cytomegalovirus β 2.7 Long Non-Coding RNA Prevents Induction of Reactive Oxygen Species to Maintain Viral Gene Silencing during Latency. *International Journal of Molecular Sciences*, 2022, 23(19): 11017.
- [41] Schmerer N, Schulte LN. Long noncoding RNAs in bacterial infection. *WIREs RNA*, 2021, 12(6): e1664.
- [42] Cheng Y, Liang Y, Tan X, et al. Host long noncoding RNAs in bacterial infections. *Frontiers in Immunology*, 2024(15).
- [43] Sun H, Ke C, Zhang L, et al. Long Non-Coding RNA (LncRNA)-ATB Promotes Inflammation, Cell Apoptosis and Senescence in Transforming Growth Factor- β 1 (TGF- β 1) Induced Human Kidney 2 (HK-2) Cells via TGF β /SMAD2/3 Signaling Pathway. *Medical Science Monitor*, 2020, 26: e922029.
- [44] Xiao H, Zhang F, Zou Y, et al. The Function and Mechanism of Long Non-coding RNA-ATB in Cancers. *Frontiers in Physiology*, 2018, 9: 321.
- [45] Li Z, Wu X, Gu L, et al. Long non-coding RNA ATB promotes malignancy of esophageal squamous cell carcinoma by regulating miR-200b/Kindlin-2 axis. *Cell Death & Disease*, 2017, 8(6): e2888-e2888.
- [46] Simion V, Zhou H, Haemmig S, et al. A macrophage-specific lncRNA regulates apoptosis and atherosclerosis by tethering HuR in the nucleus. *Nature Communications*, 2020, 11(1): 6135.
- [47] Zhao Q, Pang G, Yang L, et al. Long Noncoding RNAs Regulate the Inflammatory Responses of Macrophages. *Cells*, 2021, 11(1).
- [48] Pan X, He L. LncRNA MEG3 expression in sepsis and its effect on LPS-induced macrophage function. *Cellular and Molecular Biology*, 2020, 66(5): 131-136.
- [49] Chen K, Shi X, Jin Y, et al. High lncRNA MEG3 expression is associated with high mortality rates in patients with sepsis and increased lipopolysaccharide-induced renal epithelial cell and cardiomyocyte apoptosis. *Experimental and Therapeutic Medicine*, 2019, 18(5): 3943-3947.
- [50] Fan H, Shao H, Gao X. Long Non-Coding RNA HOTTIP is Elevated in Patients with Sepsis and Promotes Cardiac Dysfunction. *Immunological Investigations*, 2022, 51(7): 2086-2096.
- [51] Ghafouri-Fard S, Aghabalazade A, Shoorei H, et al. The Impact of lncRNAs and miRNAs on Apoptosis in Lung Cancer. *Frontiers in Oncology*, 2021, 11: 714795.
- [52] Zhang Y, Zhang YY, Xia F, et al. Effect of lncRNA-MIAT on kidney injury in sepsis rats via regulating miR-29a expression. *European Review for Medical and Pharmacological Sciences*, 2019, 23(24): 10942-10949.
- [53] Chen W, Ruan Y, Zhao S, et al. MicroRNA-205 inhibits the apoptosis of renal tubular epithelial cells via the PTEN/Akt pathway in renal ischemia-reperfusion injury. *American Journal of Translational Research*, 2019, 11(12): 7364-7375.

- [54] Chen Y, Jing H, Tang S, et al. Non-coding RNAs in Sepsis-Associated Acute Kidney Injury. *Frontiers in Physiology*, 2022(13).
- [55] Li Z, Jin T, Yang R, et al. Long non-coding RNA PFI inhibits apoptosis of alveolar epithelial cells to alleviate lung injury via miR-328-3p/Creb1 axis. *Experimental Cell Research*, 2023, 430(1): 113685.
- [56] Yao MY, Zhang WH, Ma WT, et al. Long non-coding RNA MALAT1 exacerbates acute respiratory distress syndrome by upregulating ICAM-1 expression via microRNA-150-5p downregulation. *Aging (Albany NY)*, 2020, 12(8): 6570-6585.
- [57] Su Q, Liu Y, Lv X, et al. Inhibition of lncRNA TUG1 upregulates miR-142-3p to ameliorate myocardial injury during ischemia and reperfusion via targeting HMGB1- and Rac1-induced autophagy. *Journal of Molecular and Cellular Cardiology*, 2019, 133: 12-25.
- [58] Ding Y, Wan S, Liu W, et al. Regulation Networks of Non-Coding RNA-Associated ceRNAs in Cisplatin-Induced Acute Kidney Injury. *Cells*, 2022, 11(19): 2971.
- [59] Kay M, Soltani BM. LncRNAs in Cardiomyocyte Maturation: New Window for Cardiac Regenerative Medicine. *Non-Coding RNA*, 2021, 7(1): 20.
- [60] Xie L, Zhang Q, Mao J, et al. The Roles of lncRNA in Myocardial Infarction: Molecular Mechanisms, Diagnosis Biomarkers, and Therapeutic Perspectives. *Frontiers in Cell and Developmental Biology*, 2021, 9: 680713.
- [61] Yang Z, Lu S, Pan Y, et al. Umbilical cord mesenchymal stem cell exosomal miR-143-3p delays endothelial cell senescence through targeting COX-2. *Plos One*, 2025, 20(7): e0327173.
- [62] Chen Y, Fu Y, Song Y, et al. Increased Expression of lncRNA UCA1 and HULC Is Required for Pro-inflammatory Response During LPS Induced Sepsis in Endothelial Cells. *Frontiers in Physiology*, 2019, (10): 608.
- [63] Chen J, He Y, Zhou L, et al. Long non-coding RNA MALAT1 serves as an independent predictive biomarker for the diagnosis, severity and prognosis of patients with sepsis. *Molecular Medicine Reports*, 2020, 21(3): 1365-1373.
- [64] Lin L, Liu H, Zhang D, et al. Nanolevel Immunomodulators in Sepsis: Novel Roles, Current Perspectives, and Future Directions. *International Journal of Nanomedicine*, 2024, 19: 12529-12556.